

Effect of *Urtica dioica* agglutinin on breast cancer cell growth

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Abstract

Background: Breast cancer remains a leading cause of mortality among women globally, necessitating the exploration of novel therapies with minimal side effects. Urtica dioica agglutinin (UDA), a lectin from stinging nettle, exhibits antiproliferative properties in various cancers, but its effects on breast cancer cells remain underexplored. This study evaluates UDA's cytotoxic potential against MCF-7 breast cancer cells while assessing its impact on normal mammary (MCF-10A) and embryonic kidney (HEK-293) cells.

Methods: UDA was purified from Urtica dioica rhizomes via affinity chromatography, confirmed by SDS-PAGE (8.5–9.5 kDa) and agglutination assays. MCF-7, MCF-10A, and HEK-293 cells were treated with different concentration of UDA (7.5–480 µg/ml) for 24 and 48 hours. Cytotoxicity was assessed using MTT assays to measure cell viability.

Results: UDA significantly inhibited MCF-7 proliferation in a dose- and time-dependent manner ($P < 0.01$ at 24 hours; $P < 0.0001$ at 48 hours). At 240 µg/ml (during 48 hours), viability dropped below 50%, while normal HEK-293 cells showed <30% toxicity. MCF-10A proliferation remained unaffected, even at 480 µg/ml.

Conclusion: UDA selectively targets breast cancer cells (MCF-7) with minimal toxicity to normal cells, positioning it as a promising anticancer candidate. Further studies are needed to elucidate its mechanism of action and apoptosis-inducing potential.

Keywords: Urtica dioica agglutinin, Breast cancer, MCF-7, Cytotoxicity, Lectin

Introduction

Breast cancer is the most prevalent malignancy in women accounting for 11.7% of all global cancer cases (1). It remains a leading cause of mortality among women, with projections estimating over 1 million annual deaths by 2040 (2). In Iran, breast cancer represents approximately 28.1% of female cancers, with incidence rates expected to rise by 63% by 2025 compared to 2016 (3, 4).

Conventional therapies, including surgery, chemotherapy, and immunotherapy, face challenges such as therapy resistance and adverse side effects (5, 6). Consequently, natural products, particularly herbal medicines, have gained attention for their potential to target therapy-resistant cancer stem cells (CSCs) and reduce side effects (7).

Urtica dioica (Stinging nettle) is a perennial plant with antiproliferative and anti-oxidant properties on different cancers (8). Previous studies report its aqueous extract inhibits MCF-7 cell growth and induces apoptosis (9,10). *Urtica dioica* agglutinin (UDA), a lectin isolated from nettle rhizomes, has demonstrated inhibition of benign prostatic hyperplasia (BPH) cell proliferation (11). Lectins are carbohydrate binding proteins with special characteristics such as cell agglutination, apoptosis induction and inhibition of angiogenesis. (12). Cytotoxic effects of UDA on leukemia cell lines including Jurkat (an acute lymphoblastic leukemia cell line) (13) and HL-60 (a myeloid leukemia cell line) via remarkable apoptosis induction (14). This study evaluates UDA's cytotoxicity against MCF-7 cells while assessing its safety in normal MCF-10A and HEK-293 cells.

Methods

Purification of UDA and agglutination assay

Stinging nettles were harvested from Tonekabon, Iran, during winter and authenticated in the herbarium of Golestan Agricultural and Natural Resources Research and Education Center (herbarium number: 2541). Rhizomes were cut into small pieces after washing and used immediately or stored at -20°C. UDA was purified via affinity chromatography on a chitin column following established protocols (15). Briefly, after homogenizing rhizomes in 0.1% HCl, extraction was carried out during several stages. Total UDA was purified from *Urtica dioica* extract by affinity chromatography on column chitin equilibrated with acetate buffer. After passing the extract, unbinding proteins were washed off with 1 M NaCl. Finally, the lectin desorbed with 0.5 N acetic acid. The lyophilized UDA was dialyzed against phosphate buffer saline (PBS). The concentration of UDA was determined using the lectin-specific absorption method (16). Purity was confirmed by SDS-PAGE (8.5–9.5 kDa), and agglutination activity was tested using trypsin-treated human erythrocytes (17, 18).

Cell culture

The cell lines MCF-7 (human breast cancer cell line), MCF-10A (human cell line of normal breast epithelium) and HEK-293 (human embryonic kidney cells) were purchased from Pasteur Institute (Tehran, Iran). The cells were cultured in RPMI-1640 medium (Gibco, USA, Cot. No. 11875093) supplemented with 10% heat-inactivated fetal bovine serum (FBS) (Gibco, USA, Cot. No. A25904DG) and penicillin-streptomycin (Gibco, USA, Cot. No. 15140122).

MTT assay

Cell growth inhibition was measured based on the 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyl-tetrazoliumbromide (MTT) assay. Cells (1×10^4 cells/well) were treated with UDA (7.5–480 µg/mL) for 24 and 48 hours at 37 °C. Untreated cells served as negative control. After incubation, 20 µl of MTT solution (5 mg/ml in PBS) (Sigma, USA, A101161) was added into each well and cells were incubated at 37 °C for 4 hours. After adding DMSO to wells to dissolve the formazan

crystals, absorbance was measured at 570 nm. The experiment was performed three times. Cell viability (%) was calculated as (19):

$$\text{Cell viability (\%)} = \frac{A_{570nm}(\text{sample})}{A_{570nm}(\text{control})} \times 100$$

Results

UDA purification

Purified UDA showed agglutination activity at 20 $\mu\text{g/mL}$ and a single band (8.5–9.5 kDa) on SDS-PAGE.

Proliferation inhibition by UDA treatment

UDA inhibited MCF-7 proliferation dose- and time-dependently, with viability dropping below 50% at 240 $\mu\text{g/mL}$ (48 hours) ($P < 0.0001$). HEK-293 cells exhibited less than 30% toxicity at 480 $\mu\text{g/mL}$, while MCF-10A viability remained unaffected (Figure 1).

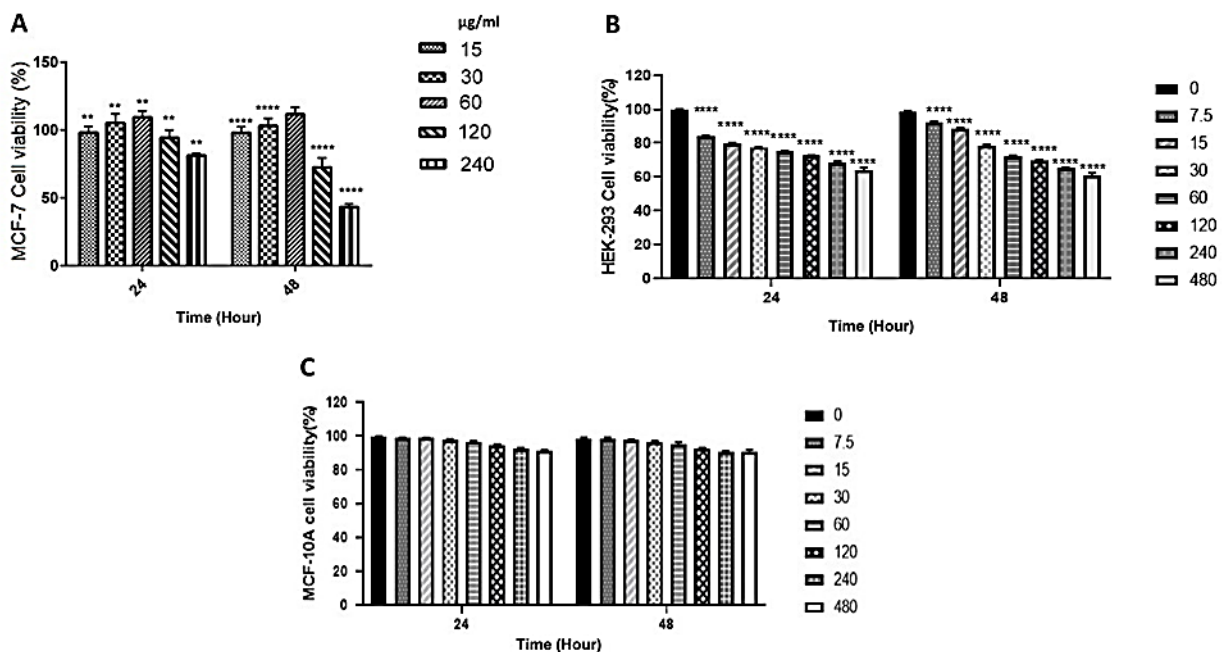


Figure 1. UDA inhibits the proliferation of MCF-7. Cell viability of (A) MCF-7, (B) MCF-10A and (C) HEK-293 cells measuring by MTT assay after 24- and 48-hour UDA treatment with different doses (15- 240 $\mu\text{g/mL}$ for MCF-7 cells and 7.5- 480 $\mu\text{g/mL}$ for MCF-10A and HEK-293 cells). Data represent mean \pm SEM of three experiments. ** $P < 0.01$, **** $P < 0.0001$.

Discussion

Here, we investigate the cytotoxic effect of UDA on a breast cancer cell line MCF-7. UDA significantly inhibited the cell growth of MCF-7 in a dose and time dependent manner ($P < 0.01$ at 24 hours and $P < 0.0001$ at 48 hours). The highest proliferation inhibition occurred at 240 $\mu\text{g/ml}$ over 48 hours, with less than 50% inhibition. However, a much weaker effect was observed in nontumorigenic human mammary MCF-10A and normal HEK293 cells.

As reported in previous studies, several lectins have showed anti-breast cancer activity (20-22). Valentin et al., 2003, examined six lectins- *Hilix pomatia* (HPA), Soybean agglutinin (SBA), Peanut agglutinin (PNA), *Solanum tuberosum* agglutinin (STA), Wheatgerm agglutinin (WGA) and Phytohemagglutinin (PHA-L)- on MCF-7 and other breast cancer cell lines. Three lectins

inhibited MCF-7 growth during a 24-hour incubation. WGA exhibited the strongest effect, reducing cell growth with an IC_{50} of 70 $\mu\text{g/ml}$. At 200 $\mu\text{g/ml}$, viable cells decreased to 28%. PHA-L showed moderate dose-dependent inhibition, reducing viability to 58% at 200 $\mu\text{g/ml}$. SBA had the weakest effect, with viable cells decreasing to 77% at the same concentration (20). The inhibitory effect of UDA was most similar to that of SBA, as UDA achieved comparable inhibition at 240 $\mu\text{g/ml}$ over 24 hours.

Deepa et al., 2012, demonstrated that mulberry leaf lectin (MLL) induced strong growth inhibition in MCF-7 cells after 24 hours (IC_{50} : 8.5 $\mu\text{g/ml}$) (). Cell cycle analysis revealed 41% of cells in the sub-G1 phase, indicating apoptosis (21). Similarly, Savanur et al. (2014) reported that *Sclerotium rolfsii* lectin (SRL) significantly inhibited MCF-7 cells. Treatment with 20 $\mu\text{g/ml}$ and 40 $\mu\text{g/ml}$ SRL reduced viability to 50% and 39%, respectively, over 48 hours. Cell cycle analysis showed 33% of treated cells in the sub-G1 phase (indicative of apoptosis) (22). While both MLL and SRL exhibited stronger inhibitory effects than UDA, further experiments are needed to confirm whether UDA induces apoptosis. If confirmed, UDA's inhibitory effect could hold significant value in breast cancer treatment.

The proliferation rates of HEK-293 and MCF-10A cells treated with UDA remained approximately 70% and >90%, respectively, at 240 $\mu\text{g/ml}$ over 48 hours. In contrast, MCF-10A cells treated with SRL retained >70% viability at a lower concentration (40 $\mu\text{g/ml}$) over the same period (22). These results suggest that UDA has fewer side effects than SRL, making it a safer candidate for therapeutic applications.

Conclusion

UDA selectively inhibits breast cancer cell proliferation with minimal impact on normal cells, supporting its potential as an anticancer agent. Future research should focus on elucidating its mechanism of action and evaluating synergistic effects with conventional therapies.

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Ethical statement

This study was conducted under the ethics approval code issued by the Golestan University of Medical Sciences (ID: IR.GOUMS.REC.1400.422).

Conflicts of interest

There is no conflict of interest to be declared by the authors.

Author contributions

All the authors made substantial, direct, and intellectual contributions to the work and read and approved the final manuscript.

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